

Valiolamine derivatives and production thereof**Patent number:** EP0199591**Publication date:** 1986-10-29**Inventor:** KAMEDA YUKIHIKO; HORII SATOSHI**Applicant:** TAKEDA CHEMICAL INDUSTRIES LTD (JP)**Classification:****- International:** A61K31/13; A61K31/70; C07C91/23; C07H15/224; C12P13/00; C12P19/46**- European:** C07H15/224C; C12P19/46; C12R1/55**Application number:** EP19860303048; 19860423**Priority number(s):** JP19850089513; 19850424**Also published as:**

US4923975 (A1)

JP62048692 (A)

EP0199591 (A3)

EP0199591 (B1)

Cited documents:

EP0089812

US4089947

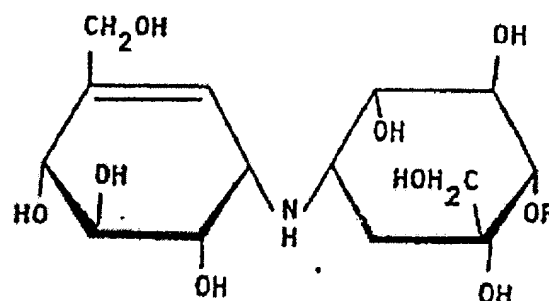
EP0049981

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Abstract not available for EP0199591

Abstract of correspondent: **US4923975**

A novel valioline derivative of the formula (I):
(I) where R stands for hydrogen or beta -D-glucopyranosyl group is useful for a starting material of valioline which can be converted to more potent alpha -glucosidase inhibiting compounds. The valioline derivative is obtained by cultivating *Streptomyces hygroscopicus* in a culture medium and recovering the valioline derivative produced and accumulated from the culture broth.



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EUROPEAN PATENT APPLICATION

②① Application number: 86303048.2

②② Date of filing: 23.04.86

⑤① Int. Cl.⁴: **C 07 H 15/224**

A 61 K 31/70, C 07 C 91/23

A 61 K 31/13, C 12 P 19/46

C 12 P 13/00

/(C12P19/46, C12R1:55),

(C12P13/00, C12R1:55)

③③ Priority: 24.04.85 JP 89513/85

④③ Date of publication of application:
29.10.86 Bulletin 86/44

⑥④ Designated Contracting States:
AT BE CH DE FR GB IT LI LU NL SE

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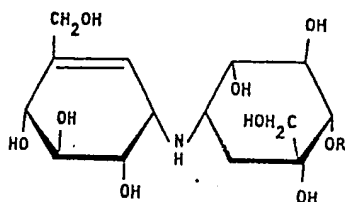
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⑤④ **Valiolamine derivatives and production thereof.**

⑥⑦ A novel valiolamine derivative of the formula (I):

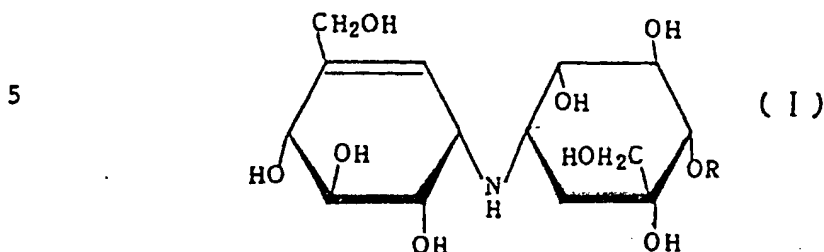


where R stands for hydrogen or a β-D-glucopyranosyl group is useful for a starting material of valiolamine which can be converted to more potent α-glucosidase inhibiting compounds. The valiolamine derivative is obtained by cultivating *Streptomyces hygroscopicus* in a culture medium and recovering the valiolamine derivative produced and accumulated from the culture broth.

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Valiolamine derivatives and production thereof

This invention relates to a novel valioline derivative of the general formula (I);



10 where R stands for hydrogen or a β-D-glucopyranosyl group and a method for production of the valioline derivative which comprises cultivating a microorganism which belongs to the genus Streptomyces and is capable of producing the valioline derivative in a culture medium
 15 to produce and accumulate the valioline derivative in the culture broth and recovering the valioline derivative from the culture broth.

The present inventors found previously that microorganisms belonging to the genus Streptomyces
 20 produced a series of validamycin antibiotics, i.e. validamycins A, B, C, D, E, and F and validoxylamines A and B [See J. Antibiotics 25, 48-53 (1972)]. The chemical structures of these compounds have been estimated as shown in Table 4 based on the results of
 25 the further studies.

The inventors have found that among the various compounds obtained by cultivation of the microorganisms under aerobic conditions there exist the novel valioline derivatives of the general formula (I) and named the compound of the formula (I) where R is hydrogen "validoxylamine G" and the compound of the formula (I) where R is a β -D-glucopyranosyl group "validamycin G". Validamycin and validoxylamine are sometimes abbreviated as VM and VA, respectively, and so validamycin G and validoxylamine G are VM-G and VA-G, respectively, in this specification.

As shown in Table 4, both VA-G and VM-G contain, as the common moieties, valioline [See J. Antibiotics 37, 1301-1307 (1984)] and valienamine [See J. Chem. Soc. Chem. Comm. 1972, 476-747] which are bound through a -NH- bond, being different from VM-A, C, D, E, and F and VA-A which contain, as the common moieties, valienamine and validamine [See J. Antibiotics 24, 59-63 (1971)] bound through a -NH- bond, and also from VM-B and VA-B which contain, as the common moieties, valienamine and hydroxyvalidamine bound through a -NH- bond [See J. Antibiotics 24, 59-63 (1971)] but not valioline.

The compounds having the general formula (I) may form various acid addition salts. For example, they may form salts of an inorganic acid such as sulfuric acid, hydrochloric acid, hydrobromic acid, phosphoric acid, nitric acid, etc. and salts of organic acid such as methanesulfonic acid, p-toluenesulfonic acid, etc. Therefore the compounds of this invention may be recovered or purified as their salts.

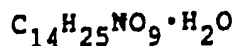
Physicochemical properties of validoxylamine G and validamycin G of this invention are as follows:

Validoxylamine G

1) Appearance

white powder

2) Elementary analysis (molecular formula)



calculated (%): C, 45.52; H, 7.36; N, 3.79

found (%): C, 45.83; H, 7.45; N, 3.62

5 3) Specific rotation

$$[\alpha]_{\text{D}}^{25} + 118.6^\circ \text{ (c=1, H}_2\text{O)}$$

4) UV absorption spectrum

The aqueous solution does not show any characteristic absorption maximum in the range 200-360 nm, except end absorption.

10 5) IR absorption spectrum

The spectrum measured by the KBr method is shown in Fig. 1. Wave numbers of main absorption peaks are shown in the following.

15 3270, 2880, 1620, 1460, 1390, 1350, 1300, 1265,
1235, 1160, 1140, 1100, 1040, 1025, 975, 930, 905,
895, 885, 860, 835 cm^{-1}

6) ^1H -NMR spectrum

Fig. 2 shows the spectrum measured in heavy water at 100 MHz (ppm; internal standard: sodium 2,2-dimethyl-2-silapentane-5-sulfonate (DSS)). The chemical shifts (δ), splitting pattern (s: singlet, d: doublet, t: triplet, dd: double doublet, dt: double triplet, ABq: AB quartet, m: multiplet) and coupling constant (J) were δ 1.50 (1H, dd, J=3 Hz, 15.5 Hz, H-6), 2.12 (1H, dd, J=3 Hz, 15.5 Hz, H-6), 3.36-4.20 (12H), and 6.06 (1H, d, J=4.3 Hz, H-2'). Table 1 shows the chemical shifts (δ), splitting pattern and coupling constant (J) of validoxylamine octaacetate obtained by acetylation of validoxylamine G with acetic anhydride-pyridine, in CDCl_3 at 400 MHz (ppm; internal standard: tetramethylsilan (TMS)).

7) ^{13}C -NMR spectrum

The chemical shifts (δ) under decoupling

conditions in heavy water at 100 MHz (ppm; internal standard: DSS) and splitting pattern under off-resonance conditions are shown in Table 2.

8) Solubility in solvents

- 5 Soluble in water, dimethylsulfoxide, and methanol.
Slightly soluble or insoluble in ethanol, ethyl acetate, chloroform, and acetone.

9) Color reaction

- Reaction with Greig-Leaback reagent: positive
10 Naphthoresorcinol-sulfuric acid reaction: negative

10) Thin layer chromatography

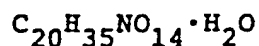
- Rf values measured on TLC plate: silica gel
60F₂₅₄ (produced by Merck & Co.) developed with
1-PrOH-AcOH-H₂O (4:1:1) (developing solvent I) or
15 1-BuOH-MeOH-CHCl₃-28%NH₄OH (4:5:2:5) (developing solvent II) are listed in Table 3.

Validamycin G

1) Appearance

white powder

- 20 2) Elementary analysis (molecular formula)



calculated (%) : C, 45.19; H, 7.01; N, 2.63

found (%) : C, 45.31; H, 7.19; N, 2.47

3) Specific rotation

- 25 $[\alpha]_D^{25} + 52.8^\circ$ (c=1, H₂O)

4) UV absorption spectrum

The aqueous solution does not show any characteristic absorption maximum in the range 200-360 nm, except end absorption.

- 30 5) IR absorption spectrum

The spectrum measured by the KBr method is shown in Fig. 3. Wave numbers of main absorption peaks are shown in the following.

- 3370, 2900, 1635, 1410, 1260, 1160, 1075, 1030,
35 900, 845 cm⁻¹

6) ^1H -NMR spectrum

Shown in Fig. 4 are the spectral data obtained in heavy water at 100 MHz (ppm; internal standard: DSS). The chemical shifts (δ), splitting pattern, and coupling constant (J) were δ 1.53 (1H, dd, J=3 Hz, 15.5 Hz, H-6), 2.12 (1H, dd, J=3 Hz, 15.5 Hz, H-6), 3.30-4.20 (18H), 4.47 (1H, d, J=7.1 Hz, H-1"), and 6.04 (1H, d, J=4.6 Hz, H-2'). Chemical shift (δ), splitting pattern, and coupling constant (J) of validamycin G undecaacetate obtained by acetylation of validamycin G with acetic anhydride-pyridine, measured in CDCl_3 at 400 MHz (ppm; internal standard: TMS) are shown in Table 1.

7) ^{13}C -NMR spectrum

Values of Chemical shifts (δ) under decoupling conditions in heavy water at 100 MHz (ppm; internal standard: DSS) and splitting pattern under off-resonance conditions are shown in Table 2.

8) Solubility in solvents

Soluble in water, dimethylsulfoxide and methanol. Slightly soluble or insoluble in ethanol, ethyl acetate, chloroform, and acetone.

9) Color reaction

Reaction with Greig-Leaback reagent: positive

Naphthoresorcinol-sulfuric acid reaction: positive

10) Thin layer chromatography

Rf values on TLC plate: silica gel 60F₂₅₄ (produced by Merck & Co.) developed with 1-PrOH-AcOH-H₂O (4:1:1) (developing solvent I) or 1-BuOH-MeOH-CHCl₃-28%NH₄OH (4:5:2:5) (developing solvent II) are listed in Table 3.

Table 1 ^1H NMR (400 MHz) data of Validoxylamine G octaacetate and Validamycin G undecaacetate in CDCl_3

Validoxylamine G octaacetate		
Proton	Chemical shifts δ^*	Splitting Pattern and Coupling Constant J(Hz)
H-1	3.547	dt $J=2.9, 3.1, 4.6$
H-2	5.018	dd $J=4.6, 10.3$
H-3	5.613	t $J=10.3$
H-4	5.069	d $J=10.3$
H-6	1.688	dd $J=2.9, 15.4$
	1.942	dd $J=3.1, 15.4$
H-7	3.653	ABq $J=11.4$
	4.042	
H-1'	3.603	m
H-2'	5.995	d $J=5.1$
H-4'	5.401	d $J=6.8$
H-5'	5.435	dd $J=6.8, 10.6$
H-6'	4.983	dd $J=5.1, 10.6$
H-7'	4.384	ABq $J=13.4$
	4.601	
NH	6.594	s
COCH_3	2.017	s
	2.048	s
	2.067	s
	2.068	s
	2.071×2	s
	2.078	s
	2.139	s

* Internal standard : TMS

Validamycin G undecaacetate		
Proton	Chemical shifts δ^*	Splitting pattern Coupling Constant J (Hz)
H-1	3.426	dt J=2.7, 3.1, 4.4
H-2	4.928	dd J=4.4, 9.9
H-3	5.522	t J=9.9,
H-4	3.665	d J=9.9,
H-6	1.492	dd J=2.7, 15.8
	1.895	dd J=3.1, 15.3
H-7	3.922	ABq J=11.1
	4.127	
H-1'	3.598	m
H-2'	5.973	d J=4.6
H-4'	5.383	d J=6.6
H-5'	5.417	dd J=6.6, 10.2
H-6'	4.985	dd J=5.1, 10.2
H-7'	4.371	ABq J=13.3
	4.599	
H-1''	4.498	d J=9.1
H-2''	4.967	t J=9.1
H-3''	5.163	t J=9.1
H-4''	5.089	t J=9.1
H-5''	3.632	ddd J=2.3, 4.1, 9.1
H-6''	4.037	dd J=2.3, 12.5
	4.404	dd J=4.1, 12.5
NH	5.870	br s
COCH ₃	1.989	s 2.061 s
	2.007	s 2.065 s
	2.032	s 2.083 s
	2.044	s 2.102 s
	2.052	s 2.141 s
	2.058	s

* Internal standard: TMS

Table 2 ^{13}C NMR Spectrum of Validoxylamine G
and Validamycin G

Chemical shifts data δ (ppm) *		
	Validoxylamine G	Validamycin G
C - 1	56.7 (d)	56.4 (d)
C - 2	75.5 (d)	75.4 (d)
C - 3	74.3 (d)	72.9 (d)
C - 4	76.6 (d)	86.0 (d)
C - 5	78.4 (s)	79.0 (s)
C - 6	31.4 (t)	31.8 (t)
C - 7	67.8 (t)	67.1 (t)
C - 1'	54.4 (d)	54.5 (d)
C - 2'	125.3 (d)	125.0 (d)
C - 3'	142.2 (s)	142.1 (s)
C - 4'	75.6 (d)	75.8 (d)
C - 5'	74.3 (d)	74.3 (d)
C - 6'	72.4 (d)	72.5 (d)
C - 7'	64.3 (t)	64.3 (t)
C - 1''		105.8 (d)
C - 2''		76.2 (d)
C - 3''		78.5 (d)
C - 4''		72.1 (d)
C - 5''		78.7 (d)
C - 6''		63.2 (t)

*Internal standard : DSS

Table 3 Rf values on thin layer chromatography of
Validamycin group antibiotics

Compound	I	II
Validoxylamine A	0.30	0.40
B	0.37	0.82
G	0.23	0.24
Validamycin A	0.20	0.29
B	0.29	0.23
C	0.14	0.14
D	0.20	0.21
E	0.14	0.21
F	0.14	0.21
G	0.18	0.14

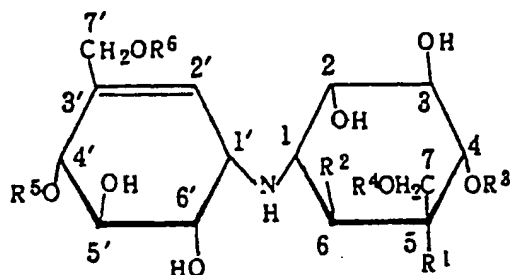
Solvent system I : 1-PrOH-AcOH-H₂O (4:1:1)

II : 1-BuOH-MeOH-CHCl₃-28%

NH₄OH (4:5:2:5)

Thin layer silica gel 60F₂₅₄ (Merck)

Table 4



- VA-A $R^1 = R^2 = R^3 = R^4 = R^5 = R^6 = H$
 VA-B $R^2 = OH, R^1 = R^3 = R^4 = R^5 = R^6 = H$
 VA-G $R^1 = OH, R^2 = R^3 = R^4 = R^5 = R^6 = H$
 VM-A $R^3 = \beta\text{-D-Glc}, R^1 = R^2 = R^4 = R^5 = R^6 = H$
 VM-B $R^2 = OH, R^1 = R^5 = R^6 = H,$
 $\beta\text{-D-Glc}$ is attached to one of the hydroxyl groups
 in the hydroxy validamine moiety, but the hydroxyl group
 which $\beta\text{-D-Glc}$ is attached to has not been specified yet.
 VM-C $R^3 = \beta\text{-D-Glc}, R^6 = \alpha\text{-D-Glc}, R^1 = R^2 =$
 $R^4 = R^5 = H$
 VM-D $R^4 = \alpha\text{-D-Glc}, R^1 = R^2 = R^3 = R^5 = R^6 = H$
 VM-E $R^3 = \alpha\text{-D-Glc}-(1-4)\text{-}\beta\text{-D-Glc}, R^1 =$
 $R^2 = R^4 = R^5 = R^6 = H$
 VM-F $R^3 = \beta\text{-D-Glc}, R^5 = \alpha\text{-D-Glc}, R^1 = R^2 =$
 $R^4 = R^6 = H,$
 VM-G $R^1 = OH, R^3 = \beta\text{-D-Glc}, R^2 = R^4 = R^5 =$
 $R^6 = H$

VA = Validoxylamine, VM = Valicamycin,
 VE = Valienamine, VD = Validamine,
 $\alpha\text{-D-Glc} = \alpha\text{-D-glucopyranosyl},$
 $\beta\text{-D-Glc} = \beta\text{-D-glucopyranosyl},$

Validoxylamine G and validamycin G of this invention are novel compounds which have not been disclosed in any literature yet, and are produced, for example, by the following method; Streptomyces hygrosopicus var. limoneus belonging to the genus Streptomyces, is cultivated under aerobic conditions to produce validoxylamine G and validamycin G.

Streptomyces hygrosopicus var. limoneus is deposited as IFO 12703 at the Institute for Fermentation, Osaka, as FERM-P No. 468 at the Fermentation Research Institute, Agency of Industrial Science and Technology, and as ATCC 21431 at The American Type Culture Collection.

As one of the common properties of microorganisms, the strain described above also can mutate spontaneously or in response to a mutagen. Mutants obtained by radiation with radiant rays such as X-rays, γ -rays, UV-rays, etc., by single spore isolation, by treatment with various chemicals, by cultivation in the medium containing a chemical, or by other methods, and mutants occurring spontaneously, which can produce validamycin G and/or validoxylamine G may all be utilized for the method of this invention.

The media used for cultivation of the microorganisms described above in this invention may be liquid or solid as far as they contain nutrients which the strain may utilize, but liquid media are preferred on a large scale cultivation. The media are suitably admixed with the carbon and nitrogen sources that the microorganisms can assimilate and digest, inorganic substances, trace nutrients, etc. As the carbon source, for example, glucose, lactose, sucrose, maltose, dextrin, starch, glycerin, mannitol, sorbitol, and oils and fats (e.g. soybean oil, lard oil, chicken oil, etc.); as the nitrogen source, meat extract, yeast extract, dried yeast,

powdered soybean, corn steep liquor, peptone, cotton seed flour, spent molasses, urea, ammonium salts (e.g. ammonium sulfate, ammonium chloride, ammonium nitrate, ammonium acetate, etc.) and others are used. In
5 addition, metal salts such as salts of sodium, potassium, calcium, magnesium, iron, manganese, zinc, cobalt and nickel, salts of inorganic acids such as phosphoric acid and boric acid, and salts of organic acids such as acetic acid and propionic acid are used appropriately. Further,
10 amino acids (e.g. glutamic acid, aspartic acid, alanine, valine, lysine, methionine, proline, etc.), vitamins (e.g. B₁, B₂, nicotinic acid, B₁₂, C, etc.) and nucleic acids (e.g. purine and pyrimidine, and their derivatives, etc.) may be contained. Inorganic or
15 organic acids, alkali, buffer, etc. may be added to adjust the pH of the media, and suitable amounts of oils and fats, surfactants, etc. may be added as defoaming agents.

Means of cultivation include standing culture,
20 shaken culture, and submerged culture. It is needless to say that the submerged culture is desirable for a large scale cultivation. Conditions of cultivation are of course variable depending upon the state and composition of the medium, the strain, means
25 of cultivation, etc., but it is advisable to select the temperature of 20-45°C and initial pH in the range of about 6-8. Preferably the temperature at the middle stage of cultivation is 24-37°C with the initial pH of 6.5-8.5. The cultivation period is also variable according
30 to the conditions described above, but it is advisable to cultivate until the concentrations of validoxylamine G and/or validamycin G attain their maxima. The period required for this is usually about 24-192 hours, preferably 72-168 hours by shaken culture or submerged
35 culture with the use of liquid media.

For the recovery of the objective substances from the culture broth, means used usually for recovery of metabolites of microorganisms are employed solely or in combination thereof in an arbitrary order or repeatedly.

- 5 That is, for example, filtration, centrifugation, concentration, drying, freeze-drying, adsorption, desorption, procedures utilizing the difference in solubilities in various solvents (e.g. precipitation, crystallization, etc.), chromatography, etc. are used.
- 10 In addition, by taking advantage of the fact that validoxylamine G and validamycin G are water-soluble basic substances, use is advantageously made of the procedure employed for isolation and purification of so-called water-soluble basic substances, e.g.
- 15 chromatography and adsorption-desorption using ion-exchange resin, activated carbon, high porous polymer, cephadex, cephadex ion-exchanger, cellulose, ion-exchange cellulose, silica gel, alumina, etc.

- More specifically speaking, the objective products may be obtained for example by the following processes:
- 20 mycelia are removed from the culture broth by centrifugation or by filtration, followed by adsorption of the objective substances onto a chromatographic column of activated carbon desirably after the pH of the supernatant or of the filtrate is made basic, and then
- 25 elution with an aqueous solution containing lower alcohols such as methanol, ethanol, n-propyl alcohol and isopropyl alcohol, n-butyl alcohol and acetone. The objective substances may be adsorbed onto a
- 30 cation-exchange resin (e.g. an ion-exchange resin containing sulfonic acid functional group such as Dowex 50W x 8 etc.) and eluted with acid (e.g. hydrochloric acid), alkali (e.g. aqueous ammonia solution of sodium hydroxide), or buffer (e.g. pyridine-acetic acid buffer).
- 35 The resultant eluate is concentrated under reduced

pressure, and the objective substances may be purified, if necessary, by activated carbon chromatography repeatedly. The objective substances may be purified and isolated by chromatography on an anion exchange resin
5 such as Dowex 1 x 2 (OH⁻ form) followed by elution with, for example, water.

The compounds of this invention, VA-G and VM-G, have, like other validamycin antibiotics, an activity against the sheath blight of rice plant. For example,
10 the minimum concentration causing the abnormal branching at the tops of hypha of *Rhizoctonia solanii* [See J. Antibiotics 24, 114-118 (1971)] was 0.5 µg/mL for VM-G and 2.5 µg/mL for VA-G.

In addition VA-G and VM-G have also
15 α-glucosidase-inhibiting activity, for example, IC₅₀ (molarity required for 50% inhibition of the enzymatic activity when the substrate is 50 mM sucrose) against sucrase from porcine small intestine was 1.1×10^{-4} M for VM-G and 8.8×10^{-6} M for VA-G. Therefore, these
20 compounds may be used as drugs for various diseases due to hyperglycemia, for example as anti-obesity drugs or as anti-diabetic drugs.

Furthermore, VA-G and VM-G are useful compounds as materials for the production of valioline. That is,
25 valioline may be produced, for example, by chemical degradation of VA-G and VM-G or enzymatic degradation using enzymes of microorganisms. Methods of chemical degradation include those which give valioline by hydrogenolysis of VA-G in the presence of a catalyst for
30 catalytic reduction, such as platinum dioxide, platinum black, palladium black, palladium-carbon, and Raney nickel, in a solvent such as water, acetic acid-water, methanol-water, and ethanol-water. The reaction is usually performed at room temperature under atmospheric
35 pressure, but may be performed under heating or

- 15 -

pressure. When VM-G is a starting material, the objective valioline may be obtained, for example, by hydrogenolysis of VM-G, as in the case of VA-G described above, followed by hydrolysis of the resultant

5 β -D-glucopyranosylvaliolamine.

Methods of enzymatic degradation include the treatment of VM-G or VA-G with the culture broth or the treated culture broth of microorganisms that can produce valioline from VM-G or VA-G, e.g. Flavobacterium
10 saccharophilum (IFO 13984, FERM-P No. 5707), Cytophaga
heparina (IFO 12017), Agrobacterium radiobacter (IFO 12664) and Agrobacterium tumefaciens (IFO 3058). "The treated culture broth" described above means bacterial cells or enzyme-containing ground cells obtained by
15 physico-chemical treatment of the culture, e.g. filtration, centrifugation, ultrasonication, French press treatment, grinding with alumina, treatment with bacteriolytic enzymes, treatment with surfactant, treatment with organic solvent, etc.; enzymes obtained
20 by known purification by prior art methods of purification such as filtration, centrifugation, ammonium sulfate fractionation, dialysis, chromatography, electrophoresis, and gel filtration of the ground cells; and the cells or enzymes immobilized
25 by known methods.

Valiolamine is a potent α -glucosidase-inhibitor and in addition it is an important compound as the starting material for more potent
 α -glucosidase-inhibitors, N-substituted valioline
30 derivatives (European Patent Publication No. 56194 and 89812).

The N-substituted valioline derivatives having potent α -glucosidase-inhibitory action include:

- (1) N-phenethylvaliolamine,
- 35 (2) N-(3-phenylallyl)valiolamine,

- (3) N-furfurylvaliolamine,
(4) N-thenylvaliolamine,
(5) N-(3-pyridylmethyl)valiolamine,
(6) N-(4-bromobenzyl)valiolamine,
5 (7) N-[(R)- β -hydroxyphenethyl]valiolamine,
(8) N-[(S)- β -hydroxyphenethyl]valiolamine,
(9) N-(β -hydroxy-2-methoxyphenethyl)valiolamine,
(10) N-(3,5-di-tert-butyl-4-hydroxybenzyl)valiolamine,
(11) N-(cyclohexylmethyl)valiolamine,
10 (12) N-geranylvaliolamine,
(13) N-(1,3-dihydroxy-2-propyl)valiolamine,
(14) N-(1,3-dihydroxy-1-phenyl-2-propyl)valiolamine,
(15) N-[(R)- α -(hydroxymethyl)benzyl]valiolamine,
(16) N-cyclohexylvaliolamine,
15 (17) N-(2-hydroxycyclohexyl)valiolamine,
(18) N-[(1R,2R)-2-hydroxycyclohexyl]valiolamine,
(19) N-(2-hydroxycyclopentyl)valiolamine,
(20) Methyl 4-[(1S,2S)-(2,4,5(OH)/3,5)-2,3,4,5-
tetrahydroxy-5-(hydroxymethyl)cyclohexyl]amino-
20 4,6-dideoxy- α -D-glucopyranoside,
(21) Methyl 4-[(1S,2S)-(2,4,5(OH)/3,5)-2,3,4,5-
tetrahydroxy-5-(hydroxymethyl)cyclohexyl]amino-4-
deoxy- α -D-glucopyranoside,
(22) [(1S,2S)-(2,4,5(OH)/3,5)-2,3,4,5-tetrahydroxy-5-
25 (hydroxymethyl)cyclohexyl][(1R,2S)-(2,6/3,4)-4-
amino-2,3-dihydroxy-6-(hydroxymethyl)cyclohexyl]-
amine,
(23) N-[(1R,2S)-(2,4/3,5)-2,3,4-trihydroxy-5-
(hydroxymethyl)cyclohexyl]valiolamine,
30 (24) N-[(1R,2S)-(2,6/3,4)-4-amino-2,3-dihydroxy-6-
methylcyclohexyl]valiolamine,
(25) N-[(1R,2S)-(2,6/3,4)-2,3,4-trihydroxy-6-
methylcyclohexyl]valiolamine,
(26) N-[(1R,2S)-(2,4,6/3)-2,3,4-trihydroxy-6-
35 methylcyclohexyl]valiolamine,

- (27) 4-O- α -[4-[(1S)-(1,2,4,5 (OH)/3,5)-2,3,4,5-tetrahydroxy-5-(hydroxymethyl)cyclohexyl]amino]-4,6-dideoxy-D-glucopyranosyl]-D-glucopyranose,
- (28) 1,6-anhydro-4-O- α -[4-[(1S)-(1,2,4,5 (OH)/3,5)-2,3,4,5-tetrahydroxy-5-(hydroxymethyl)cyclohexyl]amino]-4,6-dideoxy-D-glucopyranosyl]- β -D-glucopyranose.

The invention is illustrated by reference examples and examples.

10 Reference Example 1

Validoxylamine G (100 mg) was dissolved in 0.05 M phosphate buffer (pH 7.0) (20 mL) and wet cells of Flavobacterium saccharophilum IFO 13984 (about 2 g) were added to the solution, followed by incubation at 27°C for 24 hours with stirring. The reaction mixture was centrifuged for removal of cells, and the supernatant was adsorbed onto a column (50 mL) of Amberlite IRC-50 (NH₄⁺ form, manufactured by Rohm & Haas Co.), followed by washing with water and elution with 0.5N aqueous ammonia. The eluate was concentrated to dryness under reduced pressure, and the residue was adsorbed again onto a column (50 mL) of Amberlite IRC-50 (NH₄⁺ form) and eluted with 0.1N aqueous ammonia. In succession to the fractions containing valienamine (eluate fractions 120-140 mL), the fractions containing valioline were eluted (eluate fractions 340-430 mL). The fractions containing valioline were concentrated to dryness under reduced pressure, and the residue was submitted to column chromatography on Dowex 1 x 2 (OH⁻ form, manufactured by Dow Chemicals Co.; 2 mL) followed by concentration of the eluate to dryness under reduced pressure, to give a white powder of valioline (35 mg).

Reference Example 2

In a similar manner to that in Reference Example 1, validamycin G (150 mg) was submitted to enzymatic

degradation using wet cells of Flavobacterium
saccharophilum IFO 13984 (about 2 g), to give a white
powder of valioline (34 mg).

Reference Example 3

5 An aqueous solution (50 mL) of validoxylamine G
(420 mg) was submitted to catalytic reduction at room
temperature under atmospheric pressure in the presence
of platinum oxide (20 mg). The catalyst was removed by
filtration, and the filtrate was adsorbed onto a column
10 of Amberlite CG-50 (NH_4^+ form, 50 mL) followed by
washing with water and then elution with 0.5N aqueous
ammonia (200 mL). The eluate was concentrated under
reduced pressure, and submitted to column chromatography
on Dowex 1 x 2 (OH^- form, 100 mL) and eluted with
15 water. The eluate fractions (120-200 mL) were
concentrated to dryness under reduced pressure, to give a
white powder of valioline (60 mg).

Reference Example 4

20 An aqueous solution (30 mL) of validamycin G (106
mg) was submitted to catalytic reduction at room
temperature under atmospheric pressure in the presence
of platinum oxide (10 mg). The catalyst was removed by
filtration and the filtrate was adsorbed onto a column
of Amberlite CG-50 (NH_4^+ form, 30 mL) followed by
25 washing with water and then elution with 0.5N aqueous
ammonia. The eluate was concentrated to dryness under
reduced pressure, and the residue was dissolved in 1N
sulfuric acid (20 mL) and hydrolyzed at 80°C for 10
hours. The reaction mixture was neutralized with barium
30 hydroxide and the supernatant obtained by centrifugation
was concentrated under reduced pressure. The
concentrate was adsorbed onto a column of Amberlite
CG-50 (ammonium form, 20 mL) followed by washing with
water and then elution with 0.5N aqueous ammonia. The
35 eluate was concentrated under reduced pressure and

submitted to column chromatography on Dowex 1 x 2 (OH⁻ form, 100 mL) followed by elution with water. The eluate fractions were concentrated under reduced pressure and then freeze-dried, to give a white powder of
5 valioline (11 mg).

Example 1

- a) To the medium (glucose 3%, soybean flour 2.2%, peptone 0.3%, calcium carbonate 0.4%, water 500 mL, pH 7) in a sterilized 2L Sakaguchi flask, a loopful of
10 Streptomyces hygrosopicus var. limoneus (IFO 12703, Ferm 468, ATCC 21431) cultivated on glucose-asparagine agar medium was inoculated and incubated at 28°C for 48 hours on a rotary shaker. Five hundred mL of the culture was transferred to the medium (glucose 5%,
15 soybean flour 3.6%, peptone 0.5%, calcium carbonate 0.4%, water 30 L, pH 7) in a sterilized 50 L stainless steel fermentation tank and incubated at 28°C for 114 hours by submerged culture. The resultant culture was filtered after pH adjustment to 9 and addition of a filter
20 aid, and the filtrate (25 L) was adsorbed onto a column (10 L) of activated carbon followed by washing with water and elution with 7% n-butyl alcohol-water. The eluate was concentrated to dryness under reduced pressure, to give 123 g of a crude powder.
- 25 b) One hundred grams of the crude powder containing validamycin G and validoxylamine G obtained in Example 1a) was submitted to column chromatography on Dowex 1 x 2 (OH⁻ form, manufactured by Dow Chemicals; 1.8L), eluted with water to give validoxylamine G-
30 containing fractions (eluate fractions 3-5 L; abbreviated as 'eluate fraction I' hereafter) and validamycin G-containing fractions (eluate fractions 6-8 L; abbreviated as 'eluate fraction II' hereafter). The eluate fraction I and the eluate fraction II were
35 separately concentrated to dryness to give 2.7 and 3.0 g

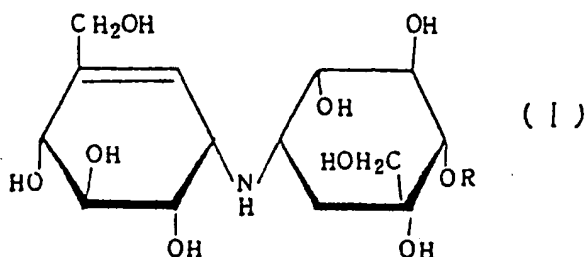
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of a crude powder, respectively. The crude powder from the eluate fraction I (2.7 g) was submitted to column chromatography on Dowex 50W x 8 (pyridine form, manufactured by Dow Chemicals Co.; 200 mL) and eluted with 0.2 M pyridine-acetate buffer (pH 6.0). In succession to the fractions containing validoxylamine B and validamycin D (eluate fractions 100-420 mL), validoxylamine G-containing fractions (eluate fractions 520-1140 mL) were eluted. The validoxylamine G-containing eluate fractions were concentrated to dryness under reduced pressure, to give a white powder (0.8 g) of validoxylamine G.

The crude powder from the eluate fraction II (3.0 g) was submitted to column chromatography on Dowex 50W x 8 (pyridine form, 200 mL) and eluted with 0.2 M pyridine-acetate buffer (pH 6.0). In succession to the validamycin C-containing fractions (eluate fractions 120-220 mL) validamycin G-containing fractions (eluate fractions 460-620 mL) were eluted. The validamycin G-containing fractions were concentrated to dryness under reduced pressure, and the residue was submitted to column chromatography on Dowex 1 x 2 (OH⁻ form, 50 mL) and eluted with water. The validamycin G-containing fractions were concentrated to dryness under reduced pressure, to give a white powder of validamycin G (0.6 g).

What is claimed is:

1. A valioline derivative of the general formula (I):

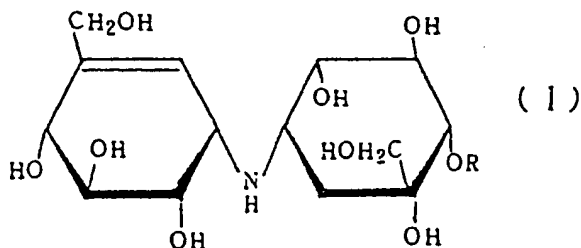


where R stands for hydrogen or a β -D-glucopyranosyl group.

2. A valioline derivative according to claim 1, wherein R stands for hydrogen, namely, validoxylamine G.

3. A valioline derivative according to claim 1, wherein R stands for a β -D-glucopyranosyl group, namely, validamycin G.

4. A method for production of a valioline derivative of the general formula (I):



where R stands for hydrogen or a β -D-glucopyranosyl group which comprises cultivating in a culture medium a microorganism which belongs to the genus Streptomyces and is capable of producing the valioline derivative to produce and accumulate the valioline derivative in the culture broth and recovering the valioline derivative from the culture broth.

5. A method for production of the valioline derivative according to claim 4, wherein the microorganism is Streptomyces hygroscopicus.

6. A method for production of the valioline

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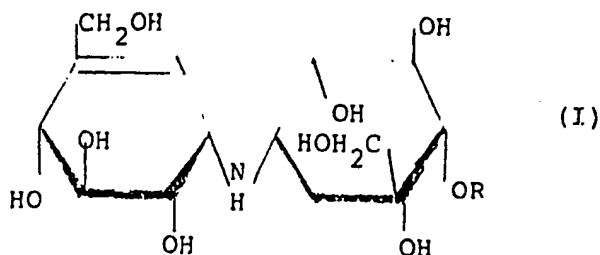
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derivative according to claim 4, wherein the
microorganism is Streptomyces hygrosopicus var.
limomeus.

Austrian Claims

What is claimed is:

1. A method for production of a valioline derivative of the general formula (I):



where R stands for hydrogen or a β -D-glucopyranosyl group which comprises cultivating in a culture medium a microorganism which belongs to the genus Streptomyces and is capable of producing the valioline derivative to produce and accumulate the valioline derivative in the culture broth and recovering the valioline derivative from the culture broth.

2. A method for production of a valioline derivative according to claim 1, wherein R stands for hydrogen, namely, validoxylamine G.
3. A method for production of a valioline derivative according to claim 1, wherein R stands for a β -D-glucopyranosyl group, namely, validamycin G.
4. A method for production of the valioline derivative according to claim 1, wherein the microorganism is Streptomyces hygroscopicus.
5. A method for production of the valioline derivative according to claim 1, wherein the microorganism is Streptomyces hygroscopicus var. limomeus.

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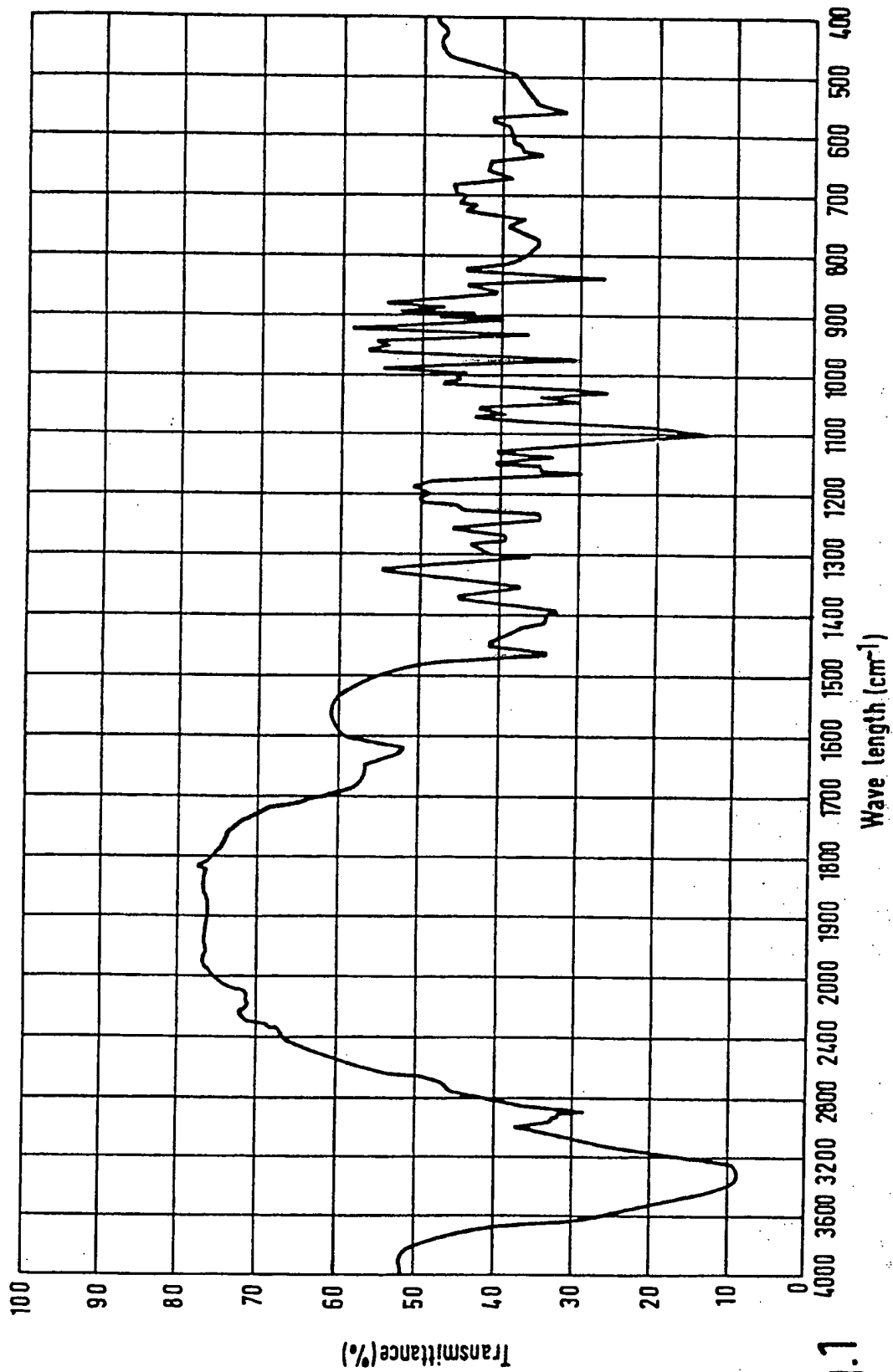
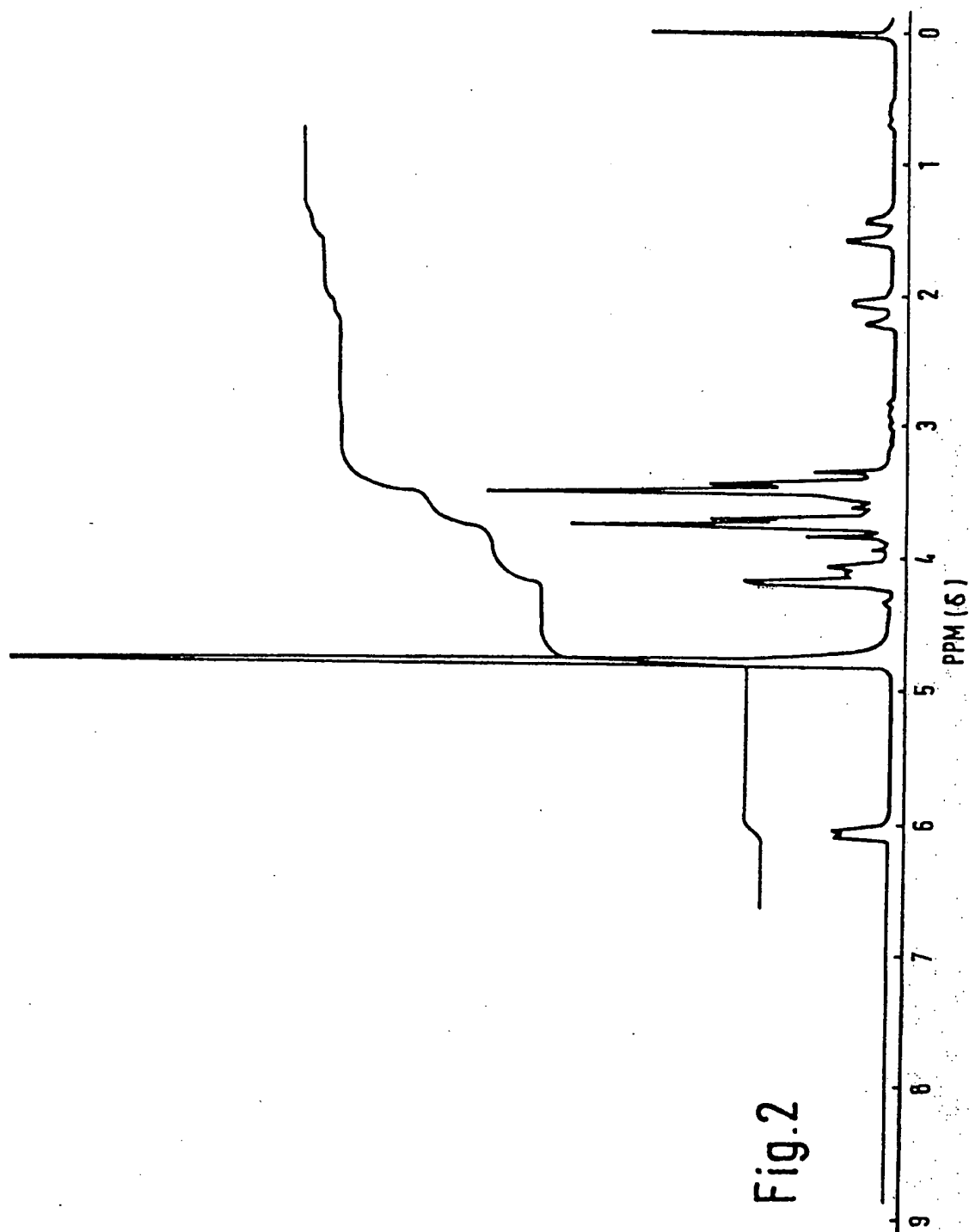


Fig.1



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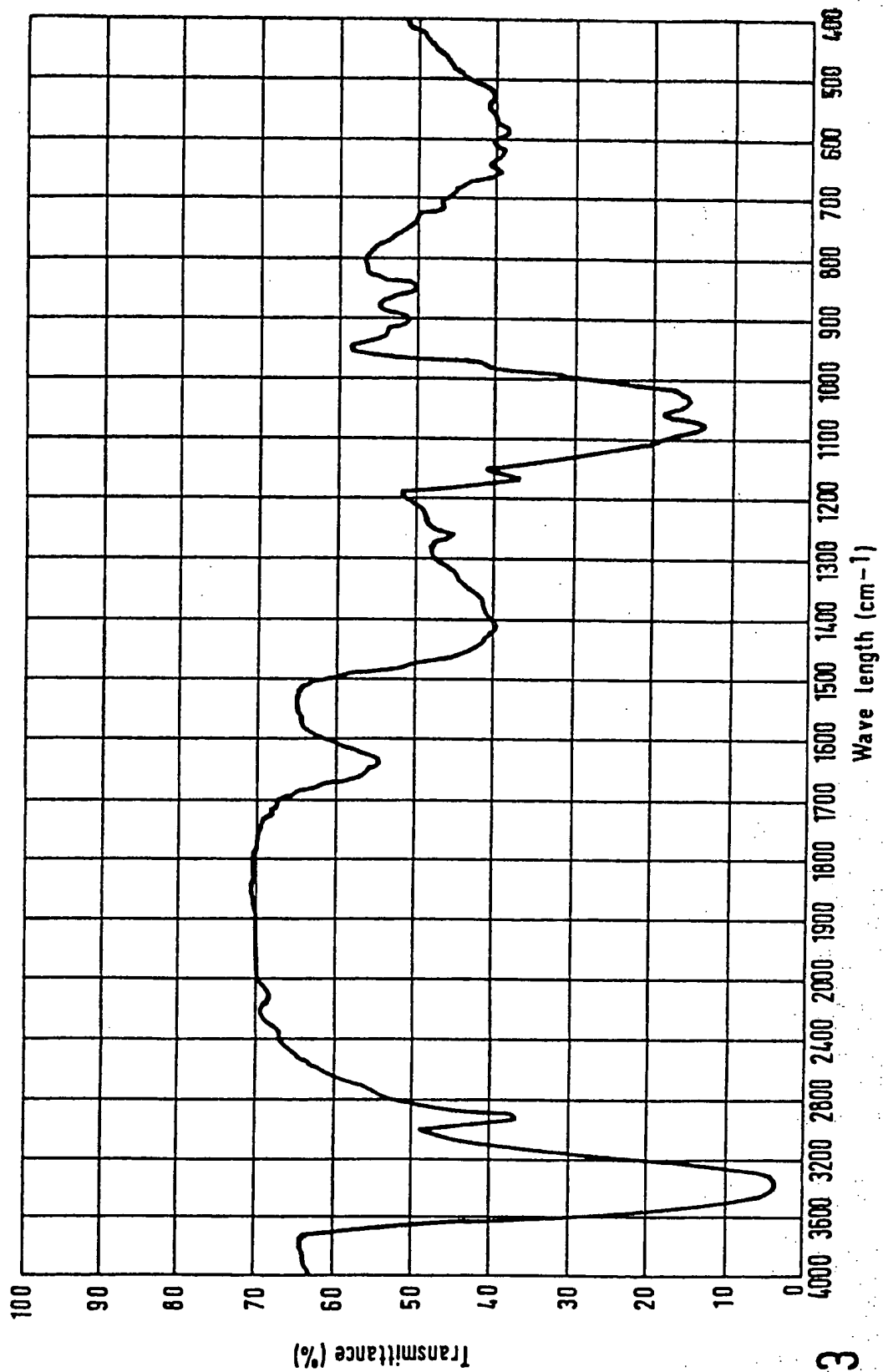


Fig.3

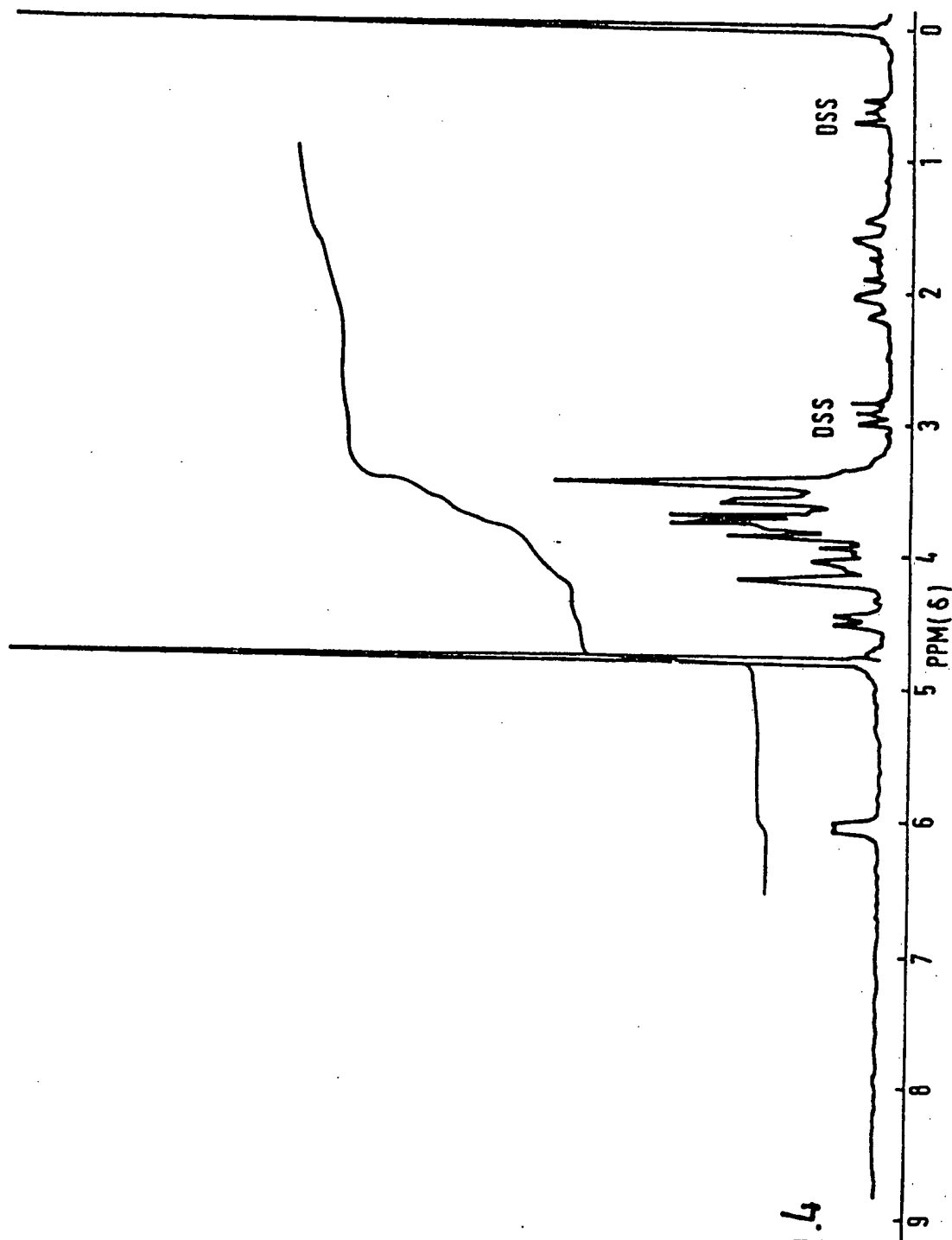


Fig. 4